Palladium-Catalyzed Ring Expansion of Spirocyclopropanes to Form Caprolactams and Azepanes

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S Supporting Information

ABSTRACT: A palladium(0)-catalyzed rearrangement of piperidones and piperidines bearing a spirocyclopropane ring was developed. The ring expansion reaction led to a variety of functionalized caprolactam and azepane products in good to excellent yields. Experimental and computational mechanistic studies revealed an initial oxidative addition of the distal carbon—carbon bond of a cyclopropane ring to the palladium(0) catalyst and the relief of ring strain as a driving force for product formation.

INTRODUCTION

Seven-membered nitrogen-containing heterocycles are prominently featured in biogically active natural products¹ and marketed drugs² and play a critical role in material science.³ Despite the prevalence of this structural motif, slow cyclization kinetics have hindered the development of robust methods for the direct construction of these medium-sized heterocycles.⁴ At present, these systems are more commonly accessed via a select few ringexpansion processes, such as migratory shift reactions, cleavage of a shared bond in fused [4.1.0] or [3.2.0] bicyclic systems, and electrocyclization reactions.⁵ Further exploration of novel ring-expansion pathways leading to seven-membered nitrogencontaining heterocycles is therefore of interest.

Because of their unique structural and electronic properties, cyclopropanes may undergo transition metal-promoted cleavage of the otherwise inert carbon-carbon bond.^{6,7} We envisaged that disubstituted cyclopropane 1 (Figure 1A) could undergo C-C bond cleavage in the presence of a transition metal via one of two postulated mechanistic pathways, both leading to common ring-open π -allyl intermediate 3. Path A would involve an initial oxidative addition of the C-X bond of cyclopropane 1 to transition metal species [M], giving rise to metalated intermediate 2. Subsequent C–C bond cleavage via β -carbon elimination⁸ would afford π -allyl intermediate 3. Alternatively, in path B, direct oxidative addition of the distal C-C bond of cyclopropane 1 to the metal would lead to metallacycle intermediate 4. This intermediate could then undergo rearrangement to π -allyl intermediate 3 through β -elimination of X. In cases where X and Y form part of a spirocyclic piperidone or piperidine ring system, the resulting π -allyl intermediate would represent a ring expanded metallacycle poised to undergo reductive elimination to the corresponding caprolactam or azepane, respectively.

Herein, we report the palladium(0)-catalyzed rearrangement of δ -lactams and saturated N-heterocycles bearing a spirocyclopropane ring, which furnished the corresponding 7-membered heterocycles (Figure 1B). In addition, the ring-expanded products featured a unique δ -exomethylene functionality primed for further



Pd(0) cat.

Ph

Pd(I

Figure 1. Mechanistic pathways for the transition metal-promoted C-C bond cleavage of cyclopropane **1** and Pd(0)-catalyzed rearrangement of lactams and piperidines bearing a spirocyclopropane ring.

late-stage diversification. Experimental and computational mechanistic studies were also performed to elucidate the mechanistic pathway of this transformation. To our knowledge, this is the first reported palladium(0)-catalyzed C–C bond cleavage reaction of a spirocyclic cyclopropane that led to further functionalized ringexpanded products.^{9–11}

RESULTS AND DISCUSSION

Optimization and Scope. Initial reaction development and optimization on the Pd(0)-catalyzed rearrangement of lactam 5 using $Pd(OAc)_2$ as a precatalyst revealed the superiority of electron-rich biarylphosphine ligands (Table 1).¹² Although MePhos (Table 1, entry 1) resulted in low conversion to

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Table 1. Optimization of the Reaction Conditions

| | Bn N - | Pd(OAc) ₂ (5 mo ligand (10 mol ⁴ base (1.5 equi solvent (0.3 M 110 °C, 16 h | $ \overset{(\%)}{\overset{(\%)}{\overset{(\gamma)}{}}} Bn_{N} \overset{(\gamma)}{\overset{(\gamma)}{}} $ | 5 |
|-----------------|--|---|--|-------------------------|
| entry | ligand | base | solvent | conversion ^a |
| 1 | MePhos | Cs ₂ CO ₃ | DMF | 12 |
| 2 | DavePhos | Cs ₂ CO ₃ | DMF | 79 |
| 3 | RuPhos | Cs ₂ CO ₃ | DMF | 97 |
| 4 | BINAP | Cs ₂ CO ₃ | DMF | 2 |
| 5 | PPh ₃ | Cs_2CO_3 | DMF | 0 |
| 6 | RuPhos | no base | DMF | 38 |
| 7 | RuPhos | NaOt-Bu | DMF | 0 |
| 8 | RuPhos | Cs ₂ CO ₃ | toluene | 94 |
| 9 | RuPhos | Cs ₂ CO ₃ | t-AmOH | 100 |
| 10 ^b | RuPhos | Cs ₂ CO ₃ | t-AmOH | 0 |
| - | 11 | 1 | • • | , h |



rearranged caprolactam 6 (12%), as determined by ¹H NMR analysis of the crude reaction mixture, significant improvements were achieved with DavePhos (entry 2) and RuPhos (entry 3), leading to conversions of 79 and 97%, respectively. Conversely, bidentate ligand BINAP (entry 4) and triphenylphospine (entry 5) resulted in only trace amounts or even no observed formation of product 6.

Although not necessary for the generatation of product 6, the addition of a base proved critical to achieve complete conversion. For example, although the combination of $Pd(OAc)_2$ and RuPhos in conjunction with Cs2CO3 gave rise to almost complete conversion (entry 3), the same reaction in the absence of base provided rearranged product 6 in a more modest 38% conversion (entry 6). From these data, it can be inferred that the addition of excess carbonate favored the formation of a stabilized $Pd(0)L_{n}X^{-}$ anionic complex.¹³ This may represent the resting state of the catalyst and would explain the increased observed turnover in the presence of carbonate. Partial decomplexation to a more active $Pd(0)L_{n-1}X^{-}$ complex presumably gave rise to the active catalyst, which underwent oxidative insertion. Conversely, in the presence of a strong yet sterically hindered base such as NaOt-Bu (entry 7), the reaction was completely suppressed with no product formation observed, highlighting the need for a more coordinating base for the reaction to proceed efficiently.¹

Several solvents proved to be tolerated in this reaction with high conversion also achieved with toluene and DMF (entries 3 and 8, respectively). However, *tert*-amyl alcohol proved to be optimal and furnished the rearranged product with complete conversion in the most consistent manner (entry 9). In the absence of palladium, the reaction did not proceed and complete recovery of δ -lactam 5 was observed (entry 10).

Having established optimized conditions for the rearrangement of N-benzyllactam 5, we evaluated the applicability of the transformation using different substituted δ -lactams, as well as *oxo*-lactams and *aza*-lactams. As illustrated in Table 2, different N-alkyl substituents were tolerated, and N-benzyllactam 6 and N-methyllactam 7 were isolated in 82 and 75% yields, respectively. Substitution at the lactam nitrogen was not required for the Pd(0)-catalyzed ring expansion reaction to proceed smoothly, as exemplified by the synthesis of NH-lactam 8 in 65% isolated yield. N-arylated subtrates possessing a broad range of electronic properties also gave rise to the rearranged caprolactam





^{*a*}All reactions were performed on 40 mg scale. ^{*b*}Reaction ran for 20 min.

products in good to excellent yields. For example, both N-phenyllactam 9 and electron-poor N-(4-nitrophenyl)lactam 10 were obtained in 74%, and electron-rich N-(4-dimethyl-aminophenyl)lactam 11 was isolated in 99% yield. Furthermore, *oxo*-lactam 12 was also compatible with the reaction conditions and was delivered in 73% yield. Similarly, a Boc-protected *aza*-lactam successfully provided desired heterocyclic product 13 in 56%. However, basic amines were not tolerated, and N-benzyl*aza*-lactam 14 was not formed under the reaction conditions.

N-Substituted piperidines also proved to be efficient substrates for the Pd(0)-catalyzed rearrangement reaction (Table 3). For example, N-benzoylpiperidine was effectively converted to the corresponding azepane 15 in 70% yield. Sulfonamide and urea functionalities were also compatible and resulted in the formation of products 16 and 17 in 76 and 94% yields, respectively. Lastly, homomorpholine 18 and homopiperazine 19 were successfully obtained from the corresponding morpholine and piperazine starting materials in 88 and 48% yields, respectively. Overall, a broad range of δ -lactams and saturated N-heterocycles reacted to yield the rearranged products, and the only observed limitation was the incompatibility of basic amines. It can therefore be inferred that the highly Lewis basic nature of these systems inhibited the catalyst by occupying an open coordination site required for the reaction to proceed, rendering it inactive toward the desired transformation.¹⁵

Mechanistic Studies. There is limited available information pertaining to the mechanism of the palladium-catalyzed C-C bond cleavage of unactivated cyclopropanes. Quantum mechanical studies reported by Blomberg¹⁶ revealed that the C-C bond of unsubstituted cyclopropane was easier to break when compared to ethane and cyclobutane. This was due to a

Table 3. Scope of N-Substituted Azepanes and Other Saturated N-Heterocycles^{*a*}



^aAll reactions were performed on 40 mg scale.

low activation barrier for the elimination reaction of metallacyclobutane, which in turn led to a small energy barrier for the reverse C–C bond oxidative addition process rather than depending strictly on the innate C–C bond strength.

Activated methylenecyclopropane C-C bond cleavage was also studied in the context of trimethylenemethane cycloaddition reactions.¹⁷ The mechanism proposed by Trost¹⁸ does not invoke palladium-mediated C-C bond oxidative addition. Conversely, Binger¹⁹ proposed a mechanism including a palladacyclobutane intermediate. Subsequent theoretical studies by Fujimoto²⁰ supported the involvement of a palladacyclobutane intermediate, but no unifying mechanism had yet been established. Yamamoto²¹ also demonstrated that methylenecyclopropanes may undergo ring opening via a palladium-catalyzed C-Cbond oxidative addition to form palladacyclobutane intermediates in the presence of carbon-based nucleophiles. However, in the presence of nitrogen nucleophiles, C-C bond cleavage was proposed to proceed through β -carbon elimination.²² More recently, Fu^9 reported the Pd(0)-catalyzed C-C bond cleavage of electronically activated 1,1-difluoro-2-arylcyclopropanes. Preliminary DFT calculations demonstrated that initial oxidative addition of a cyclopropane C-C bond to form a palladacyclobutane intermediate followed by β -fluorine elimination (Figure 1A, path B) was lower in energy than an initial C-F oxidative addition followed by β -carbon elimination (Figure 1A, Path A).

Similarly, we applied computational methods to investigate the two proposed mechanistic pathways for the cyclopropane ring expansion reaction of lactam **20** illustrated in Figure 2. All calculations were performed using DFT with the B3LYP density functional. The LANL2DZ basis set with the Hay–Wadt effective core potential was used for the Pd atom.²³ For all other atoms, the 6-31* basis set was used. All stationary points were confirmed to be minima or transition states by computing the frequencies on optimized structures.²⁴

Because of the more extensive computational power required for large systems, few reported all-atom DFT studies have been performed using the full structure of biarylphosphine



Figure 2. Computationally calculated mechanistic pathways for the rearrangement of piperidone **20** to caprolactam 7. (a) Energies are reported as the sum of the energy of the separated compounds **20** or 7 and catalyst **27**.

ligands,²⁵ such as RuPhos,²⁶ which performed best in our reaction. However, it has been shown that potentially significant differences in calculated energies may occur when approximating the structure of ligands.²⁷ With this consideration in mind, we decided to elect biphenyldimethylphosphine (see Figure 2, complex 27) as our model ligand for further DFT studies. We conducted initial optimization on the active monoligated LPd(0) catalyst.²⁸ The lowest energy conformation of the catalyst displayed an η^1 interaction between the biphenyl substituent of the ligand and the palladium(0) center. We then performed geometry optimizations to identify low energy conformations of oxidative addition products 22 and 24 on the two pathways shown in Figure 2.²⁹ Optimizations were initiated from multiple input geometries to identify the lowest energy conformations.²⁴ The corresponding transition states 21 and 23 were identified using the QST2 method³⁰ in Gaussian.³¹ The oxidative addition of the C–N bond of lactam 20 (Figure 2, path A) to palladium catalyst 27 was computed to present a transition state energy barrier of +32.8 kcal/mol (TS 21), yielding palladacycle 22 at +8 kcal/mol. Conversely, it was revealed that the addition of the distal C-C bond of the cyclopropane ring (Figure 2, path B) had a much lower transition state energy of +3.9 kcal/mol (TS 23), giving rise to palladacyclobutane 24 at -6.9 kcal/mol. On the basis of these results, it was concluded that path B, which involved the formation of a palladacyclobutane intermediate, was the operative reaction mechanism.³²

It was hypothesized that the relief of the cyclopropane ring strain was the driving force that allowed for the low activation

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energy barrier at the C–C bond cleavage step in TS 23 and the generation of energetically downhill palladacyclobutane intermediate 24. In contrast, the high energy barrier associated with the formation of palladacycle 22 in path A was hypothesized to be the result of a more sterically encumbered C–N bond precluding the approach of the active palladium center during the oxidative addition step. Additionally, intermediate 22 did not benefit from the relief of cyclopropane ring strain.

Both palladacycle intermediates **22** and **24** could then converge to common π -allyl complex **25** via a β -carbon elimination (Figure 2, path A) or a β -nitrogen elimination (Figure 2, path B), respectively. It was calculated that π -allyl intermediate **25** possessed an energy of -6.6 kcal, similar to palladacyclobutane **24**. Subsequent reductive elimination via TS **26** was judged to be rate-determining with an energy barrier of +18.9 kcal/mol leading to product 7 (-5.6 kcal/mol).³³

To further analyze our proposed mechanism, we attempted to experimentally trap the various palladacycle intermediates under hydrogenative conditions and isolate the hydrogenated adducts for characterization (Figure 3). Treatment of lactam **28**



Figure 3. Treatment of lactams $\mathbf{28}$ and $\mathbf{9}$ under hydrogenation conditions.

under our optimized reaction conditions, with the addition of 1 atm of hydrogen, resulted in the formation of hydrogenated adduct 30 in 15% isolated yield. Other isolated products were the ring expanded product (9, 45% yield) and the hydrogenated olefin product (49, 35% yield, see Scheme 6). It was hypothesized that the formation of acyclic amide 30 was the product of homogeneous hydrogenation of π -allyl complex 29. This result confirmed the presence of this key π -allyl intermediate in the reaction mixture. Additionally, subjecting caprolactam 9 to the same reductive reaction conditions gave rise to the same amount of hydrogenated product (30). The generation of 30 from both cyclopropylpiperidone 28 and caprolactam 9 demonstrated that all reaction intermediates and products were in equilibrium under the reaction conditions. This was in agreement with the computed rate-determining energy barrier relative to the penultimate π -allyl intermediate (-6.6 kcal/mol) and the final product (-5.6 kcal/mol) energies. Interestingly, we did not observe a gem-dimethyl product, which would have resulted from the homogeneous hydrogenation of the calculated low-energy palladacyclobutane intermediate. Because both the palladacyclobutane (not shown) and the π -allyl complex (29) were calculated to possess similar ground state energies (-6.9 and -6.6 kcal/mol, respectively), it was hypothesized that both species should also be in equilibrium and in at a similar concentration in the reaction mixture. By applying the Curtin-Hammett principle,³⁴ it can be inferred that the corresponding barrier that produced hydrogenated amide product 30 from π -allyl complex 29 was lower in energy than the energy

barrier that would lead to a gem-dimethyl product from the calculated palladacyclobutane intermediate. As such, only 30 formed in the reaction mixture.

CONCLUSIONS

In conclusion, we established Pd(0)-catalyzed conditions for the ring opening of spirocyclopropanes that led to the formation of functionalized caprolactams and azepanes in good to excellent yields. Mechanistic studies uncovered a low-energy palladacyclobutane intermediate that was energetically favored due to the relief of ring strain as a driving force. This represents the first Pd(0)-catalyzed C–C bond cleavage reaction of a spirocyclic cyclopropane that led to functionalized medium-sized nitrogencontaining heterocycles. In addition to its synthetic utility, this process also demonstrated the viability of cyclopropanes as reactive coupling partners in transition-metal-catalyzed synthetic transformations.

EXPERIMENTAL SECTION

General Information. Reaction mixtures were analyzed on a UPLC-MS system using formic acid/MeCN mobile phases and a C18 column (1.7 μ m, 2.1 \times 30 mm). ¹H and ¹³C NMR spectra were recorded in CDCl₃ solutions at 400 MHz for ¹H and 101 MHz for ¹³C. The internal standard is TMS (0.00 ppm) for the resonance of protons and residual chloroform (77.0 ppm) for the resonance of carbons. High resolution mass spectra were obtained by positive EI and orbitrap mass analysis. Flash column chromatography purification was performed using disposable normal-phase 15–40 μ m silica gel columns. All reaction solvents were purchased anhydrous from commercial sources and used as is. The following compounds were purchased from commercial sources and were used as is **31** [546114-04-9], **32** [1100753-07-8], **33** [1199794-51-8], **39** [1199794-52-9], **41** [1301739-56-9], **42** [218595-22-3], **43** [886766-28-5].³⁵

General Procedure for Tables 2 and 3: Ring Expansion Reactions. A sealable reaction vial was charged with the spirocyclopropane starting material (40 mg), palladium(II) acetate (0.05 equiv), RuPhos (0.10 equiv), and cesium carbonate (1.5 equiv) and then evacuated under vacuum and backfilled with argon (3×). *tert*-Amyl alcohol (0.3M) was added, and the resulting suspension was stirred and heated at 110 °C for 16 h. The reaction was then washed with CH_2Cl_2 (3 × 5 mL) and filtered through a pad of Celite. Concentrating the resulting filtrate followed by silica gel column chromatography (0–60% acetone in heptane) afforded the corresponding caprolactam or azepane product.

1-Benzyl-6-methylene-azepan-2-one (6). Yield: 33 mg, 82%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 7.27 (m, 5H), 4.81 (s, 1H), 4.62 (m, 3H), 3.72 (s, 2H), 2.69 (m, 2H), 2.36 (t, *J* = 6.0 Hz, 2H), 1.83(m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 174.8, 143.3, 137.5, 128.5, 128.2, 127.3, 113.9, 53.2, 50.1, 37.0, 36.1, 24.0; HRMS calcd for C₁₄H₁₇NO [M + H]⁺ 216.1383, found 216.1391.

1-Methyl-6-methylene-azepan-2-one (7). Yield: 30 mg, 75%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 4.88 (m, 2H), 3.80 (d, J = 0.7 Hz, 2H), 2.99 (s, 3H), 2.61(m, 2H), 2.37 (m, 2H), 1.78 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 174.7, 143.4, 113.6, 56.0, 36.9, 35.8, 34.8, 24.0; HRMS calcd for C₈H₁₃NO [M + H]⁺ 140.1070, found 140.1075.

6-Methyleneazepan-2-one (8). Yield: 26 mg, 65%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 6.28 (s, 1H), 4.88 (d, *J* = 12 Hz, 2H), 3.67 (d, *J* = 6.3 Hz, 2H), 2.54 (m, 2H), 2.42 (m, 2H), 1.79 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 177.6, 144.4, 114.0, 47.7, 37.4, 35.5, 23.5; HRMS calcd for C₇H₁₁NO [M + H]⁺ 126.0914, found 126.0913.

6-Methylene-1-phenyl-azepan-2-one (9). Yield: 29.5 mg, 74%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 7.36 (m, 2H), 7.22 (m, 3H), 4.96 (s, 1H), 4.83(s, 1H), 4.20 (s, 2H), 2.78 (m, 2H), 2.49 (m, 2H), 1.92 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 174.3, 144.0, 143.8, 129.1, 126.7, 126.6, 114.2, 57.5, 36.7, 36.4, 23.7; HRMS calcd for C₁₃H₁₅NO [M + H]⁺ 202.1227, found 202.1233.

6-Methylene-1-(4-nitrophenyl)azepan-2-one (**10**). Reaction was run for 20 min instead of 16 h. Yield: 29.5 mg, 74%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 8.23 (d, *J* = 9.1 Hz, 2H), 7.44 (d, *J* = 8.9 Hz, 2H), 5.07–5.01 (m, 1H), 4.95–4.88 (m, 1H), 4.29 (s, 2H), 2.86–2.77 (m, 2H), 2.51 (td, *J* = 6.2, 1.1 Hz, 2H), 2.01–1.86 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 174.2, 149.4, 145.4, 143.2, 126.7, 124.4, 114.7, 57.0, 36.4, 36.2, 23.3; HRMS calcd for C₁₃H₁₄N₂O₃ [M + H]⁺ 247.1077, found 247.1093.

1-(4-(Dimethylamino)phenyl)-6-methyleneazepan-2-one (11). Yield: 39.5 mg, >95%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 7.10–6.97 (m, 2H), 6.78–6.63 (m, 2H), 4.98–4.90 (m, 1H), 4.87–4.79 (m, 1H), 4.15 (s, 2H), 2.94 (s, 6H), 2.80–2.71 (m, 2H), 2.52–2.44 (m, 2H), 1.97–1.85 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 174.6, 149.3, 144.0, 133.5, 127.1, 113.9, 112.9, 57.9, 40.7, 36.9, 36.4, 23.8; HRMS calcd for C₁₅H₂₀N₂O [M + H]⁺ 245.1648, found 245.1667.

4-Benzyl-6-methylene-1,4-oxazepan-3-one (12). Yield: 29 mg, 73%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 7.29 (m, 5H), 5.07 (s, 1H), 4.93 (s, 1H), 4.66 (s, 2H), 4.32 (s, 2H), 4.23(s, 2H), 3.85(s, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 171.0, 143.0, 136.7, 128.6, 128.1, 127.5, 116.3, 73.7, 71.8, 51.6, 50.5; HRMS calcd for C₁₃H₁₅NO₂ [M + H]⁺ 218.1176, found 218.1182.

tert-Butyl 4-Isopropyl-6-methylene-3-oxo-1,4-diazepane-1-carboxylate (13). Yield: 22.5 mg, 56%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 5.15 (m, 2H), 4.86 (m, 1H), 4.13 (m, 4H), 3.66 (m, 2H), 1.46 (s, 9H), 1.13 (d, J = 6.7 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 168.7, 155.2, 142.3, 116.3, 80.8, 52.1, 51.4, 45.5, 44.9, 28.2, 19.6; HRMS calcd for C₁₄H₂₄N₂O₃ [M + H]⁺ 269.1860, found 269.1867.

(3-Methyleneazepan-1-yl)(phenyl)methanone (15). Yield: 28 mg, 70%; colorless oil; ¹H NMR (400 MHz, CDCl₃, reported a 2:1 mixture of rotamers) δ 7.46–7.31 (m, 5H), 5.00 (d, *J* = 11.3 Hz, 2/3H), 4.86 (s, 2/3H), 4.78–4.66 (m, 2/3H), 4.34 (s, 2/3H), 4.02 (s, 4/3H), 3.67–3.56 (m, 4/3H), 3.38–3.24 (m, 2/3H), 2.41–2.29 (m, 2/3H), 2.28–2.17 (m, 4/3H), 1.91–1.77 (m, 4/3H), 1.75–1.59 (m, 2H), 1.59–1.48 (m, 2/3H); ¹³C NMR (101 MHz, CDCl₃, reported as a mixture of rotamers) δ 172.0, 171.1, 147.6, 146.4, 137.0, 136.7, 129.2, 129.1, 128.3, 126.7, 126.4, 114.0, 112.3, 56.8, 52.6, 49.0, 46.5, 34.9, 34.6, 30.6, 29.8, 28.3, 28.2; HRMS calcd for C₁₄H₁₇NO [M + H]⁺ 216.1383, found 216.1388.

3-Methylene-1-(phenylsulfonyl)azepane (16). Yield: 30.5 mg, 76%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 7.84–7.78 (m, 2H), 7.61–7.47 (m, 3H), 4.90–4.86 (m, 1H), 4.84–4.81 (m, 1H), 3.94–3.87 (m, 2H), 3.23–3.16 (m, 2H), 2.32–2.24 (m, 2H), 1.79–1.69 (m, 2H), 1.64–1.53 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 146.8, 139.4, 132.3, 129.0, 126.9, 113.2, 54.6, 48.1, 34.0, 30.5, 29.3; HRMS calcd for C₁₃H₁₇NO₂S [M + H]⁺ 252.1053, found 252.1058.

N-*Methyl*-3-*methylene*-*N*-*phenylazepane*-1-*carboxamide* (17). Yield: 37.5 mg, 94%; colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 7.38–7.28 (m, 2H), 7.17–7.00 (m, 3H), 4.76 (d, *J* = 1.9 Hz, 1H), 4.70–4.62 (m, 1H), 3.65 (s, 2H), 3.26–3.12 (m, 5H), 2.24–2.07 (m, 2H), 1.67–1.57 (m, 2H), 1.57–1.48 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 161.8, 148.2, 147.2, 129.4, 124.3, 123.8, 112.3, 55.5, 48.0, 40.0, 34.4, 29.4, 28.6; HRMS calcd for C₁₅H₂₀N₂O [M + H]⁺ 245.1649, found 245.1658.

N-*Methyl*-3-*methylene*-*N*-*phenylazepane*-1-*carboxamide* (**18**). Yield: 35 mg, 88%; colorless oil; ¹H NMR (400 MHz, CDCl₃, reported as a 2:1 mixture of rotamers) δ 7.47–7.33 (m, 5H), 5.27 (br s, 1/3H), 5.11 (br s, 1/3H), 4.99 (br s, 2/3H), 4.72 (br s, 2/3H), 4.42 (br s, 2/3H), 4.29 (br s, 2/3H), 4.24 (br s, 4/3H), 4.11 (br s, 4/3H), 3.88 (br s, 2 2/3H), 3.64 (br s, 2/3H), 3.52 (br s, 2/3H); ¹³C NMR (101 MHz, CDCl₃, reported as a mixture of rotamers) δ 166.8, 140.6, 139.3, 131.5, 131.2, 124.9, 124.8, 123.8, 123.6, 122.0, 121.8, 109.9, 108.4, 69.8, 69.6, 67.8, 67.3, 50.1, 47.9, 45.5, 44.9; HRMS calcd for C₁₃H₁₆NO₂ [M + H]⁺ 218.1176, found 218.1179.

N-Methyl-3-methylene-N-phenylazepane-1-carboxamide (19). Yield: 19 mg, 48%; colorless oil; ¹H NMR (400 MHz, CDCl₃, reported as a mixture of rotamers) δ 7.49–7.29 (m, 5H), 5.38–4.47 (m, 2H), 4.29–3.79 (m, 5H), 3.71–3.25 (m, 3H), 1.46 (s, 9H); ¹³C NMR (101 MHz, CDCl₃, reported as a mixture of rotamers) δ 167.2, 167.1, 166.7, 150.3, 150.0, 139.6, 139.3, 138.3, 137.9, 137.4, 131.4, 131.3, 124.8, 123.8, 123.5, 121.9, 121.7, 110.2, 108.6, 107.1, 75.4, 75.3, 50.5, 48.0, 47.7, 47.0, 46.4, 46.1, 45.5, 44.9, 44.4, 44.0, 43.9, 43.5, 23.6; HRMS calcd for $C_{18}H_{25}N_2O_3$ [M + H]⁺ 317.1860, found 317.1863.

8-Methyl-8-azaspiro[2.5]octan-7-one (20) (Scheme 1). A roundbottomed flask was charged with 8-azaspiro[2.5]octan-7-one (200 mg,

Scheme 1. Preparation of Starting Materials 20 and 34-36



1.60 mmol) and *N,N*-dimethylformamide (5.3 mL). The resulting solution was cooled to 0 °C, and sodium hydride (60% in mineral oil, 51 mg, 1.3 mmol) was added, forming a yellow suspension. The suspension was then allowed to warm to room temperature, and iodomethane (0.13 mL, 2.1 mmol) was added. The reaction mixture was stirred for 16 h. Upon completion, the reaction was quenched with H₂O (1 mL), extracted with ethyl acetate (3 × 3 mL), and dried with anhydrous MgSO₄. Concentrating the organic layer followed by silica gel column chromatography (0–60% acetone in heptane) afforded 8-methyl-8-azaspiro[2.5]octan-7-one (64.5 mg, 29% yield) as a colorless gel. ¹H NMR (400 MHz, CDCl₃) δ 2.70 (s, 3H), 2.51 (d, *J* = 6.0 Hz, 2H), 1.86 (m, 2H), 1.69 (m, 2H), 1.01 (d, *J* = 6.0 Hz, 2H), 0.58 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 172.3, 40.3, 33.3, 32.2, 27.5, 19.3, 9.8; HRMS calcd for C₈H₁₃NO [M + H]⁺ 140.1070, found 140.1070.

8-Benzyl-8-azaspiro[2.5]octan-7-one (34) (Scheme 1). A roundbottomed flask was charged with 8-azaspiro[2.5]octan-7-one (200 mg, 1.60 mmol) and N,N-dimethylformamide (5.3 mL). The resulting solution was cooled to 0 °C, and sodium hydride (60% in mineral oil, 51 mg, 1.3 mmol) was added, forming a yellow suspension. The suspension was then allowed to warm to room temperature, and benzyl bromide (0.25 mL, 2.1 mmol) was added. The reaction mixture was stirred for 16 h. Upon completion, the reaction was quenched with H_2O (1 mL), extracted with ethyl acetate (3 × 3 mL), and dried with anhydrous MgSO4. Concentrating the organic layer followed by silica gel column chromatography (0-60% acetone in heptane) afforded 8-benzyl-8-azaspiro[2.5]octan-7-one (26.5 mg, 77% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.09 (m, 5H), 4.46 (s, 2H), 2.63 (t, J = 7.1 Hz, 2H), 1.93 (p, J = 6.9 Hz, 2H), 1.63 (t, J = 6.8 Hz, 2H), 0.93–0.85 (m, 2H), 0.60–0.52 (m, 2H); $^{13}\mathrm{C}$ NMR (101 MHz, $CDCl_3$) δ 173.3, 138.7, 128.5, 126.8, 126.7, 44.8, 39.9, 33.4, 32.2, 19.3, 11.2; HRMS calcd for $C_{14}H_{17}NO [M + H]^+$ 216.1383, found 216.1388.

8-Benzyl-5-oxa-8-azaspiro[2.5]octan-7-one (35) (Scheme 1). A round-bottomed flask was charged with 5-oxa-8-azaspiro[2.5]octan-7one (200 mg, 1.57 mmol) and N,N-dimethylformamide (3.2 mL). The resulting solution was cooled to 0 °C, and sodium hydride (60% in mineral oil, 47 mg, 1.2 mmol) was added, forming a yellow suspension. The suspension was then allowed to warm to room temperature, and benzyl bromide (0.24 mL, 2.0 mmol) was added. The reaction mixture was stirred for 16 h. Upon completion, the reaction was quenched with H_2O (1 mL), extracted with ethyl acetate (3 × 3 mL), and dried with anhydrous MgSO₄. Concentrating the organic layer followed by silica gel column chromatography (0-60% acetone in heptane) afforded 8-benzyl-8-azaspiro[2.5]octan-7-one (177 mg, 52% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 7.36-7.27 (m, 2H), 7.27-7.21 (m, 1H), 7.19 (dd, J = 7.5, 1.7 Hz, 2H), 4.45 (s, 2H), 4.43 (s, 2H), 3.69 (s, 2H), 1.02–0.95 (m, 2H), 0.71–0.65 (m, 2H); ¹³C NMR (101 MHz, $CDCl_3$) δ 168.5, 137.6, 128.7, 127.2, 126.4, 72.5, 68.2, 43.1, 39.0, 7.8; HRMS calcd for $C_{13}H_{15}NO_2$ [M + H]⁺ 218.1176, found 218.1185

tert-Butyl 8-Isopropyl-7-oxo-5,8-diazaspiro[2.5]octane-5-carboxylate (36) (Scheme 1). A round-bottomed flask was charged with tertbutyl 7-oxo-5,8-diazaspiro[2.5]octane-5-carboxylate (200 mg, 0.88 mmol) and *N*,*N*-dimethylformamide (2.9 mL). The resulting solution was cooled to 0 °C, and sodium hydride (60% in mineral oil, 53 mg, 1.3 mmol) was added, forming a yellow suspension. The suspension was then allowed to warm to room temperature, and 2-iodopropane (0.26 mL, 2.7 mmol) was added. The reaction mixture was stirred for 16 h. Upon completion, the reaction was quenched with H₂O (1 mL), extracted with ethyl acetate (3 × 3 mL), and dried with anhydrous MgSO₄. Concentrating the organic layer followed by silica gel column chromatography (0–60% acetone in heptane) afforded *tert*-butyl 8-isopropyl-7-oxo-5,8-diazaspiro[2.5]octane-5-carboxylate (85 mg, 36% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 4.20 (m, 3H), 3.23 (m, 2H), 1.46 (s, 9H), 1.26 (d, *J* = 7.0 Hz, 6H), 1.15 (d, *J* = 6.0 Hz, 2H), 0.92 (s, 2H); ¹³C NMR (101 MHz, CDCl₃, major rotamer reported) δ 170.1, 154.1, 80.3, 52.4, 49.0, 46.8, 37.1, 28.3, 20.8, 12.0; HRMS calcd for C₁₄H₂₄N₂O₃ [M + H]⁺ 269.1860, found 269.1870.

8-Phenyl-8-azaspiro[2.5]octan-7-one (28) (Scheme 2). A sealable reaction vial was charged with 8-azaspiro[2.5]octan-7-one (200 mg,

Scheme 2. Preparation of Starting Materials 28, 37, and 38



1.6 mmol), copper(I) iodide (30 mg, 0.154 mmol), and potassium phosphate (754 mg, 3.5 mmol) and then evacuated under vacuum and backfilled with argon (3×). Toluene (3.2 mL), *N*,*N*-dimethylethylenediamine (0.035 mL, 0.32 mmol), and iodobenzene (0.23 mL, 2.1 mmol) were then added, and the resulting slurry was heated at 90 °C and stirred for 16 h. The reaction was then quenched with H₂O (1 mL), diluted with EtOAc (3 × 5 mL), washed with 1 M HCl, saturated aqueous NaHCO₃, and brine, and dried with anhydrous MgSO₄. Concentrating the organic layer followed by silica gel column chromatography (0–60% acetone in heptane) afforded 8-phenyl-8-azaspiro[2.5]octan-7-one (190 mg, 59% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 7.36 (m, 2H), 7.27 (m, 1H), 7.07 (m, 2H), 2.69 (d, *J* = 8.0 Hz, 2H), 2.03 (d, *J* = 7.0 Hz, 2H), 1.88 (d, *J* = 8.0 Hz, 2H), 0.64 (s, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 172.7, 138.4, 128.9, 128.5, 127.2, 41.6, 33.1, 32.7, 19.4, 12.1; HRMS calcd for C₁₃H₁₅NO [M + H]⁺ 202.1227, found 202.1231.

8-(4-Nitrophenyl)-8-azaspiro[2.5]octan-7-one (37) (Scheme 2). A sealable reaction vial was charged with 8-azaspiro[2.5]octan-7-one (200 mg, 1.6 mmol), copper(I) iodide (30 mg, 0.154 mmol), and potassium phosphate (754 mg, 3.5 mmol) and then evacuated under vacuum and backfilled with argon (3×). Toluene (3.2 mL), N,Ndimethylethylenediamine (0.035 mL, 0.32 mmol), and 1-iodo-4nitrobenzene (522 mg, 2.1 mmol) were then added, and the resulting slurry was heated at 100 °C and stirred for 16 h. The reaction was then quenched with H_2O (1 mL), diluted with EtOAc (3 × 5 mL), washed with 1 M HCl, saturated aqueous NaHCO3, and brine, and dried with anhydrous MgSO4. Concentrating the organic layer followed by silica gel column chromatography (0-60% acetone in heptane) afforded 8-(4nitrophenyl)-8-azaspiro[2.5]octan-7-one (240 mg, 61% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 8.22 (d, J = 9.1 Hz, 2H), 7.36 (d, J = 9.1 Hz, 2H), 2.74 (t, J = 7.1 Hz, 2H), 2.14-2.03 (m, 2H), 1.98–1.88 (m, 2H), 0.91–0.83 (m, 2H), 0.74–0.66 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 173.4, 145.5, 144.7, 127.8, 124.1, 41.47, 32.9, 32.1, 19.0, 13.9; HRMS calcd for C₁₃H₁₄N₂O₃ [M + H]⁺ 247.1077, found 247.1093.

8-(4-Dimethylaminophenyl)-8-azaspiro[2.5]octan-7-one (**38**) (Scheme 2). A sealable reaction vial was charged with 8-azaspiro[2.5]-octan-7-one (200 mg, 1.6 mmol), copper(I) iodide (30 mg, 0.154 mmol), and potassium phosphate (754 mg, 3.5 mmol) and then evacuated under vacuum and backfilled with argon (3×). Toluene (3.2 mL), *N*,*N*-dimethylethylenediamine (0.035 mL, 0.32 mmol), and 4-iodo-*N*,*N*-dimethylaniline (518 mg, 2.1 mmol) were then added, and the resulting

slurry was heated at 100 °C and stirred for 20 h. The reaction was then quenched with H₂O (1 mL), diluted with EtOAc (3 × 5 mL), washed with saturated aqueous NaHCO₃ and brine, and dried with anhydrous MgSO₄. Concentrating the organic layer followed by silica gel column chromatography (0–60% acetone in heptane) afforded 8-(4-dimethylaminophenyl)-8-azaspiro[2.5]octan-7-one (148.5 mg, 38% yield) as a yellow amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 6.89 (d, *J* = 8.9 Hz, 2H), 6.68 (d, *J* = 8.9 Hz, 2H), 2.94 (s, 6H), 2.66 (t, *J* = 13.9 Hz, 2H), 2.05–1.96 (m, 2H), 1.88–1.81 (m, 2H), 0.66–0.60 (m, 2H), 0.60–0.53 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 172.7, 149.5, 129.2, 127.0, 112.6, 41.8, 40.6, 33.3, 32.7, 19.6, 11.4; HRMS calcd for C₁₅H₂₀N₂O [M + H]⁺ 245.1648, found 245.1660.

5-Benzyl-5,8-diazaspiro[2.5]octan-7-one (40) (Scheme 3). A round-bottomed flask was charged with 5,8-diazaspiro[2.5]octan-7-

Scheme 3. Preparation of Starting Material 40



one hydrochloride (200 mg, 1.59 mmol) and dichloromethane (5.3 mL), and benzaldehyde (0.16 mL, 1.59 mmol) was added. Sodium triacetoxyborohydride (531 mg, 2.38 mmol) and acetic acid (0.14 mL, 2.38 mmol) were then added, and the reaction was allowed to stir at room temperature for 16 h. The reaction was then quenched with H₂O (1 mL), diluted with EtOAc (3 × 5 mL), washed with 1 M HCl, saturated aqueous NaHCO₃, and brine, and dried with anhydrous MgSO₄. Concentrating the organic layer followed by silica gel column chromatography (0–60% acetone in heptane) afforded 5-benzyl-5,8-diazaspiro-[2.5]octan-7-one (123.5 mg, 36% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 7.30 (m, 5H), 3.62 (s, 2H), 3.25 (s, 2H), 2.53 (s, 2H), 0.75 (m, 2H), 0.65 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 170.6, 137.1, 128.8, 128.4, 127.4, 61.1, 56.3, 56.1, 34.9, 11.7; HRMS calcd for C₁₃H₁₆N₂O [M + H]⁺ 217.1336, found 217.1345.

Phenyl(4-azaspiro[2.5]octan-4-yl)methanone (44) (Scheme 4). 8-Azaspiro[2.5]octane hydrochloride (0.450 g, 3.05 mmol) and potassium

Scheme 4. Preparation of Starting Materials 44-46

| $ \begin{array}{c} \text{BzCl} \\ \text{K}_2\text{HPO}_2 \\ \text{H}_2\text{O}, \text{ rt, 15} \end{array} $ | |
|--|---------------------------------|
| 41 : Y = CH ₂ | 44 : Y = CH ₂ |
| 42 : Y = O | 45 : Y = O |
| 43 : Y = NBoc | 46: Y = NBoc |

phosphate dibasic (0.5 M in H₂O, 10 mL) were combined at room temperature. Benzoyl chloride (450 mg, 3.20 mmol) was added dropwise to the stirred solution. A precipitate formed as the reagent was added. The resulting slurry was stirred for 15 min at room temperature, and then CPME (6 mL) was added. The organic fraction was isolated, and the aqueous layer was rinsed with CPME (6 mL) a second time. The organic fractions were combined, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting oil was purified by column chromatography (25% EtOAc in heptane) to yield phenyl(4-azaspiro[2.5]-octan-4-yl)methanone as a white amorphous solid (510 mg, 74%). ¹H NMR (400 MHz, CDCl₃) δ 7.50–7.30 (m, 5H), 3.53 (br s, 2H), 1.80 (br s, 2H), 1.61 (br s, 4H), 0.58 (br s, 4H); ¹³C NMR (101 MHz, CDCl₃, reported as mixture of rotamers) δ 171.8, 169.7, 137.4, 129.6, 128.1, 127.1, 49.9, 44.8, 39.9, 38.3, 35.3, 32.2, 26.5, 24.1, 14.8; HRMS calcd for C₁₄H₁₇NO [M + H]⁺ 216.1383, found 216.1388.

Phenyl(7-oxa-4-azaspiro[2.5]octan-4-yl)methanone (45) (Scheme 4). 8-Azaspiro[2.5]octane hydrochloride (0.50 g, 3.3 mmol) and potassium phosphate dibasic (0.5 M in H₂O, 10 mL) were combined

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at room temperature. Benzoyl chloride (0.43 mL, 3.7 mmol) was added dropwise to the stirred solution. A precipitate formed as the reagent was added. The resulting slurry was stirred for 15 min at room temperature, and then CPME (6 mL) was added. The organic fraction was isolated, and the aqueous layer was rinsed with CPME (6 mL) a second time. The organic fractions were combined, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting oil was purified by column chromatography (25% EtOAc in heptane) to yield phenyl-(7-oxa-4-azaspiro[2.5]octan-4-yl)methanone as a white amorphous solid (466 mg, 58%). ¹H NMR (400 MHz, CDCl₃) δ 7.52–7.31 (m, 5H), 3.88–3.67 (m, 4H), 3.61 (s, 2H), 0.97–0.73 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) reported as mixture of rotamers) δ 170.9, 136.6, 130.2, 129.6, 128.5, 128.3, 127.5, 126.9, 73.6, 68.9, 67.3, 67.1, 48.1, 39.3, 13.6, 10.5; HRMS calcd for C₁₃H₁₆NO₂ [M + H]⁺ 218.1176, found 218.1179.

tert-Butyl 4-benzoyl-4,7-diazaspiro[2.5]octane-7-carboxylate (46) (Scheme 4). tert-Butyl 4,7-diazaspiro[2.5]octane-7-carboxylate (0.537 g, 2.5 mmol) and potassium phosphate dibasic (0.5 M in H₂O, 10 mL) were combined at room temperature. Benzoyl chloride (0.32 mL, 2.7 mmol) was added dropwise to the stirred solution. A precipitate formed as the reagent was added. The resulting slurry was stirred for 15 min at room temperature, and then CPME (6 mL) was added. The organic fraction was isolated, and the aqueous layer was rinsed with CPME (6 mL) a second time. The organic fractions were combined, dried over Na2SO4, filtered, and concentrated under reduced pressure. The resulting oil was purified by column chromatography (25% EtOAc in heptane) to yield tert-butyl 4-benzoyl-4,7-diazaspiro[2.5]octane-7-carboxylate as a white amorphous solid (720 mg, 91%). ¹H NMR (400 MHz, CDCl₃) δ 7.51-7.29 (m, 5H), 3.77-3.60 (m, 2H), 3.54-3.45 (m, 2H), 3.43 (s, 2H), 1.46 (s, 9H), 0.90-0.72 (m, 4H); ¹³C NMR (101 MHz, CDCl₃, reported as a mixture of rotamers) δ 171.0, 154.8, 136.5, 133.0, 130.1, 129.9, 128.2, 127.3, 80.0, 67.5, 51.0, 46.9, 44.1, 38.7, 28.3, 21.7, 21.2, 14.1; HRMS calcd for C₁₈H₂₅N₂O₃ [M + H]⁺ 317.1860, found 317.1863.

8-(Benzenesulfonyl)-8-azaspiro[2.5]octane (47) (Scheme 5). To a solution of 4-azaspiro[2.5]octane hydrochloride (150 mg, 1.01 mmol)



in dichloromethane (5 mL) was added triethylamine (0.43 mL, 3.04 mmol) and benzenesulfonyl chloride (0.19 mL, 1.52 mmol), and the reaction was stirred at room temperature for 16 h. The reaction was then concentrated on silica gel and purified by silica gel column chromatography (0–100% EtOAc in heptane) to give 8-(benzenesulfonyl)-8-azaspiro[2.5]octane (239 mg, 94% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 7.90–7.79 (m, 2H), 7.60–7.42 (m, 3H), 3.64–3.49 (m, 2H), 1.71–1.61 (m, 2H), 1.61–1.51 (m, 2H), 1.31–1.19 (m, 2H), 1.05–0.94 (m, 2H), 0.60–0.52 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 142.6, 132.2, 128.9, 127.1, 48.1, 39.3, 32.0, 24.3, 23.7, 13.9; HRMS calcd for C₁₃H₁₇NO₂S [M + H]⁺ 252.1053, found 252.1064.

N-Methyl-N-phenyl-8-azaspiro[2.5]octane-8-carboxamide (**48**) (Scheme 5). To a solution of 4-azaspiro[2.5]octane hydrochloride (150 mg, 1.01 mmol) in dichloromethane (5 mL) was added triethylamine (0.43 mL, 3.04 mmol) and N-methyl-N-phenylcarbamoyl chloride (258 mg, 1.52 mmol), and the reaction was stirred at room temperature for 16 h. The reaction was then concentrated on silica gel and purified by silica gel column chromatography (0–100% EtOAc in heptane) to give N-methyl-N-phenyl-8-azaspiro[2.5]octane-8-carboxamide (222 mg, 90% yield) as a white amorphous solid. ¹H NMR (400 MHz, CDCl₃) δ 7.32–7.24 (m, 2H), 7.11–6.99 (m, 3H), 3.21 (s, 3H), 3.04–2.91 (m, 2H), 1.68–1.56 (m, 2H), 1.51–1.31 (m, 4H), 0.82–0.72 (m, 2H), 0.67–0.54 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 161.0, 146.7, 129.0, 124.3, 124.1, 48.1, 38.8, 38.7, 31.6, 26.0, 24.0, 14.7; HRMS calcd for C₁₅H₂₀N₂O [M + H]⁺ 245.1649, found 245.1658.

Reaction of Piperidine 26 under Hydrogenation Conditions (Scheme 6). A sealable reaction vial was charged with 4-phenyl-4-

Scheme 6. Reaction of Piperidine 26 under Hydrogenation Conditions



azaspiro[2.5]octan-5-one (50 mg, 0.248 mmol), palladium(II) acetate (2.8 mg, 0.012 mmol), RuPhos (11.8 mg, 0.025 mmol), and cesium carbonate (121 mg, 0.37 mmol) and then evacuated under vacuum and backfilled with argon (3×). *tert*-Amyl alcohol (1 mL) was added, and the reaction mixture was evacuated under vacuum and backfilled with hydrogen (3×). A hydrogen-filled balloon was placed on top of the reaction vessel. The resulting suspension was heated at 110 °C and stirred for 16 h. The reaction was washed with CH_2Cl_2 (3 × 5 mL) and filtered through Celite. Concentrating the filtrate followed by silica gel column chromatography (0–60% acetone in heptane) afforded 5-methyl-N-phenylhex-5-enamide (7.5 mg, 15% yield) as a colorless oil, 6-methyl-1-phenylazepan-2-one (17.5 mg, 35% yield) as a colorless oil.

5-Methyl-N-phenylhex-5-enamide (**30**). ¹H NMR (400 MHz, CDCl₃) δ 7.56–7.46 (m, 2H), 7.36–7.29 (m, 2H), 7.13–7.07 (m, 1H), 4.77 (s, 1H), 4.72 (s, 1H), 2.41–2.29 (m, 2H), 2.12 (t, *J* = 7.5 Hz, 2H), 1.96–1.84 (m, 2H), 1.74 (s, 3H), 1.61 (s, 1H); ¹³C NMR (101 MHz, CDCl₃, C=O carbon could not be differentiated from baseline) δ 144.9, 137.8, 129.0, 124.2, 119.6, 110.8, 37.0, 36.9, 23.1, 22.1; HRMS calcd for C₁₃H₁₇NO [M + H]⁺ 204.1383, found 204.1388.

6-Methyl-1-phenylazepan-2-one (**49**). ¹H NMR (400 MHz, CDCl₃) δ 7.43–7.32 (m, 2H), 7.25–7.15 (m, 3H), 3.69 (dd, *J* = 15.0, 9.2 Hz, 1H), 3.46 (dt, *J* = 15.0, 1.5 Hz, 1H), 2.78–2.62 (m, 2H), 2.02–1.93 (m, 2H), 1.84–1.63 (m, 2H), 1.48–1.29 (m, 1H), 0.94 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 175.4, 144.7, 129.1, 126.4, 126.2, 59.1, 38.3, 37.5, 34.0, 22.7, 20.0; HRMS calcd for C₁₃H₁₇NO [M + H]⁺ 204.1383, found 204.1388.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b01846.

Copies of ¹H, ¹³C NMR spectra and computational methods (PDF)

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Notes

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